



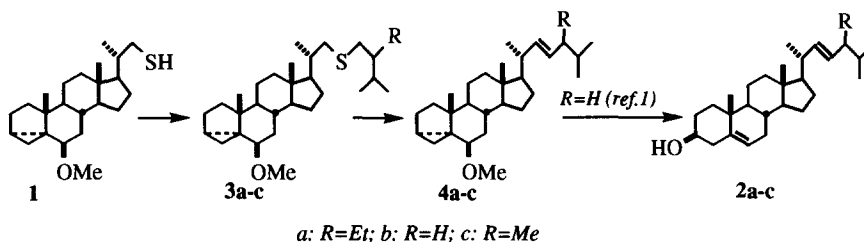
## A Formal Synthesis of Brassinolide

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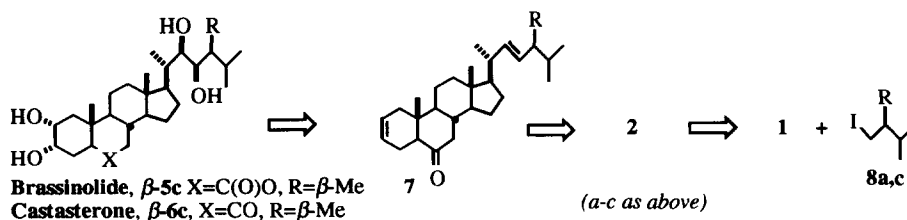
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**Abstract:** Enzyme-catalysed differentiation of hydroxy groups in a  $C_{2v}$ -shaped tetraol-sulfide, combined with a *E*-stereoconvergent Ramberg-Bäcklund process, allowed to prepare pure (2*S*)-2,3-dimethyl-1-iodobutane, which could be coupled with a 3,5-cyclopregnane-20-thiomethanol derivative so as to give an efficient precursor of the title vegetal hormone. © 1997 Published by Elsevier Science Ltd.

We have shown recently that the use of the Ramberg-Bäcklund rearrangement (RBR) allowed a convenient preparation of the side chain of sterols.<sup>1</sup> For instance, alkylation of thiol **1**, accessible in a few steps from stigmaterol -i.e.  $\beta$ -**2a**- by isoamyl iodide, gave the sulfide **3b** ( $R=H$ ), which, by a short sequence involving a chlorination, an oxidation, and treatment of the resulting chlorosulfone with excess *t*-BuOK afforded selectively the unsaturated compound **4b**, efficiently converted into  $\Delta^{22}$  dehydrocholesterol **2b**.

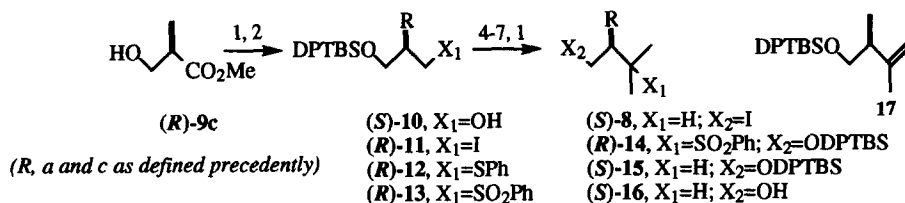


As part of our ongoing work on the synthesis of modified sterols, we planned to develop that methodology to prepare brassinolide (i.e.  $\beta$ -**5c**). This lactone is a useful plant-growth factor and various approaches have been accordingly proposed to obtain this scarcely-distributed steroid from the more accessible stigmaterol, which indeed can be converted into the ketone  $\beta$ -**7c**, then into castasterone  $\beta$ -**6c**, an immediate precursor of  $\beta$ -**5c**.<sup>2</sup> The above RBR strategy seemed apt to rival existing hemisynthetic processes but, as it appears below, implementation of such a plan necessitated prior unfolding of an efficient access to the iodide (*S*)-**8c**.



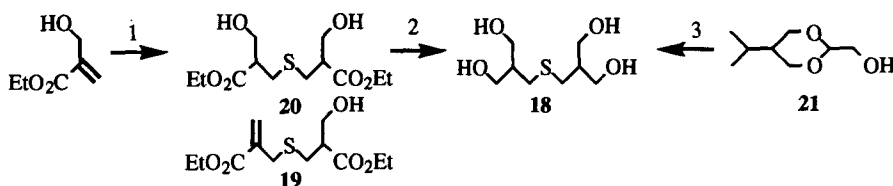
We describe herein a method allowing to prepare conveniently the iodide **8c** as well as a related 2-substituted 1-iodo-3-methylbutane (e.g. (*R*)-**8a**), which paves the way for a general access to a wide range of branched sterols.

The iodide (*S*)-**8c** (R=Me) was first prepared by a classical chiron approach, starting from commercial methyl ester of (*R*)-hydroxyisobutyric acid (*R*)-**9c** which was converted to the alcohol (*S*)-**10c**, then to the iodide (*R*)-**11c** as described previously.<sup>3</sup> Treatment of (*R*)-**11c** by PhSNa furnished the sulfide (*R*)-**12c**, which was oxidised (MCPBA) to the sulfone (*R*)-**13c**. Bis-methylation of (*R*)-**13c** (BuLi/ICH<sub>3</sub>) gave the sulfone (*R*)-**14c**, which, by hydrogenolysis of the phenylsulfonyl group (Mg/ethanol), followed by desilylation afforded the alcohol (*S*)-**16c**. Treating (*S*)-**16c** by PPh<sub>3</sub>/I<sub>2</sub> delivered the target iodide (*S*)-**8c**.



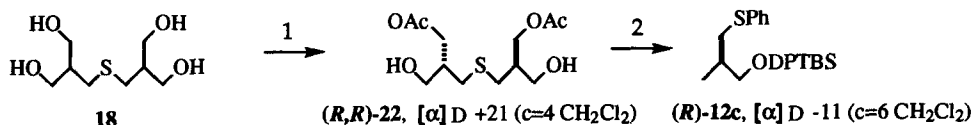
Reagents and conditions: 1- DPTBSCl (1.05 eq.), imidazole (3 eq.), DMF; r.t., 12 hours; 2- DIBA-H (2.1 eq.), CH<sub>2</sub>Cl<sub>2</sub>; -78°C, 2 hours; 3- PPh<sub>3</sub> (1 eq.), I<sub>2</sub> (1 eq.), imidazole (1 eq.); 4/1 ether/acetonitrile; r.t., 1 hour; 4- PhSNa (1.3 eq.), EtOH; r.t., 2 hours; 5- BuLi (1 eq.), ICH<sub>3</sub> (1 eq.), THF; -78°C, 0.5 hour, then same treatment (reagents added in situ), -78°C to r.t., 1 hour; 6- TBAF (1.5 eq.), THF; r.t., 5 hours; 7- Mg (5 eq.), HgCl<sub>2</sub> (cat.), EtOH; r.t. overnight.

A more enticing breakthrough resulted from the enzyme-catalysed acetylation of the C<sub>2v</sub>-shaped tetraol **18**, which was prepared, as indicated, by alumina-catalysed addition of H<sub>2</sub>S to the ethyl ester of 2-hydroxymethylacrylic acid and, after separation by column chromatography of the unsaturated sulfide **19**, which formed invariably as a side-product in that addition step, by subsequent LAH reduction of the resulting diester **20**. For larger scale preparation, an alternative, somewhat longer, procedure involving sequential treatment of the acetal **21**<sup>5</sup> by PPh<sub>3</sub>/I<sub>2</sub> and Na<sub>2</sub>S, followed by hydrolysis was preferred.



Reagents and conditions: 1- H<sub>2</sub>S, basic alumina, CH<sub>2</sub>Cl<sub>2</sub>; r.t., 12 hours (37%); 2- LAH (4 eq.), THF; r.t., 4 hours (55%); 3- i) PPh<sub>3</sub> (1 eq.), I<sub>2</sub> (1 eq.), imidazole (1 eq.); 4/1 ether/acetonitrile; 0°C-r.t., 0.5 hour (87%); ii) Na<sub>2</sub>S<sub>9</sub>H<sub>2</sub>O (0.5 eq.), EtOH; r.t., 2 days (80%); iii) 1N HCl; 110°C, with continuous distillation of volatiles (95%).

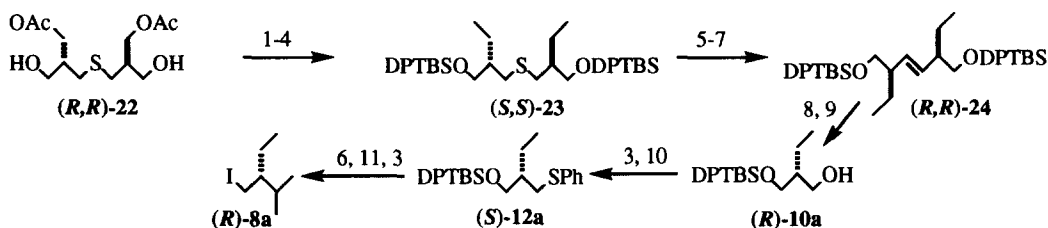
Tetraol **18** was stirred with vinyl acetate and *Pseudomonas fluorescens* lipase in THF. The acetylation process was relatively fast, compared to what had been observed previously with related polyols.<sup>6</sup> After 6 days, at what time tetraol **18** had almost (not entirely) disappeared, the acetylation process was disrupted by filtration on Celite. Evaporation of solvents, followed by column chromatography resulted in the isolation of a diacetate, which proved to be (*R,R*)-**22**, with [α]<sub>D</sub> +21 in a fairly good yield (78%). Noteworthy, any corresponding *meso* diacetate failed to be detected (<sup>13</sup>C NMR) in the crude product, which was indicative of a deserving stereoselectivity.<sup>6</sup>



Reagents and conditions: 1- PFL, vinyl acetate, THF; 5-10°C, 6 days (78%); 2- i) DPTBSCl (1.05 eq.), imidazole (3 eq.), DMF; r.t., 8 hours (93%); ii) Ni-R, EtOH; 50°C, 12 hours (93%); iii) K<sub>2</sub>CO<sub>3</sub> (2 eq.), MeOH; -10°C, 2 days (88%); iv) PPh<sub>3</sub> (1 eq.), I<sub>2</sub> (1 eq.), imidazole (1 eq.); 4/1 ether/acetonitrile; r.t., 2 hours (89%); v) PhSNa (1.3 eq.), EtOH; r.t., 2 hours (92%).

Both the nature and the level of the enantioselectivity of that acetylation process were ascertained as follows. Protection of free hydroxy groups of compound (*R,R*)-22 as DPTBS ether, then desulfuration (Raney-nickel), and hydrolysis of the acetate functionalities in mild conditions (MeOH/K<sub>2</sub>CO<sub>3</sub>) afforded the alcohol (*S*)-10, which by treatment with PPh<sub>3</sub>/I<sub>2</sub>, then with PhSNa gave the pure sulfide (*R*)-12c, identical ([α]<sub>D</sub>, NMR) to that obtained previously starting from *R*-9c and which could eventually be converted to the iodide (*S*)-8c as above.<sup>7</sup> Though lengthier to some extent than the preceding approach, that double-meso strategy proved to be of interest however for preparing a large amount of iodide (*S*)-8c since the use of the rather expensive ester (*R*)-9 was avoided.

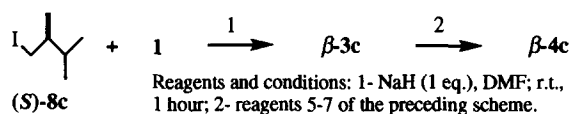
A more distinctive advantage, which is made clear in the next scheme, lies in the possibility of preparing likewise -i.e. from the same diacetate (*R,R*)-22- the iodide (*R*)-8a.



1- DPTBSCl (1.05 eq.), imidazole (3 eq.), DMF; r.t., 12 hours (92%); 2- K<sub>2</sub>CO<sub>3</sub> (2 eq.), MeOH; -10°C, 2 days (88%); 3- PPh<sub>3</sub> (1 eq.), I<sub>2</sub> (1 eq.), imidazole (1 eq.); 4/1 ether/acetonitrile; r.t., 1 hour (91%); 4- MeMgBr (2.4 eq.), Li<sub>2</sub>CuCl<sub>4</sub> (0.17 eq.), THF; -78°C- r.t., 18 hours (86%); 5- NCS (1 eq.), CCl<sub>4</sub>; 90°C, 15 mn; 6- MCPBA (3 eq.), NaHCO<sub>3</sub> (6 eq.), CH<sub>2</sub>Cl<sub>2</sub>; r.t., 3 hours; 7- *t*-BuOK (4 eq.), THF; -78°C to 0°C, 3 hours (81%); 8- O<sub>3</sub>, pyridine (0.5 eq.), DMS (1.47 eq.), CH<sub>2</sub>Cl<sub>2</sub>; -78°C; 9- DIBAH (2 eq.), CH<sub>2</sub>Cl<sub>2</sub>; -78°C, 2 hours (78%); 10- PhSNa (1.3 eq.), EtOH; r.t., 2 hours (92%); 11- *i*) BuLi (1 eq.), ICH<sub>3</sub> (1 eq.), THF; -78°C, 0.5 hour, then same treatment (reagents added in situ), -78°C to r.t., 1 hour (89%); *ii*) TBAF (1.5 eq.), THF; r.t., 5 hours (100%); *iii*) Mg (5 eq.), HgCl<sub>2</sub> (cat.), EtOH; r.t. overnight (78%).

Performing consecutively on (*R,R*)-22a a DPTBS protection, a saponification (K<sub>2</sub>CO<sub>3</sub>/MeOH), an iodination (PPh<sub>3</sub>/I<sub>2</sub>), and a methylation (MeMgBr/Li<sub>2</sub>CuCl<sub>4</sub>) afforded the bis-protected derivative (*S,S*)-23 which led to the unsaturated compound (*R,R*)-24 when submitted to RBR conditions. Ozonolysis of (*R,R*)-24, followed by DIBAH reduction provided the alcohol (*R*)-10a, which led eventually to (*R*)-8a by means of the reaction sequence used for the (*R*)-12c-(*S*)-8c conversion.

Finally, the planned synthesis of β-4c was completed by alkylating the thiol 1 by iodide (*S*)-8c and submitting the resulting sulfide β-3c to *E*-selective RBR conditions, which afforded the pure (<sup>13</sup>C NMR) crinostene derivative β-4c in a satisfactory 83% yield.



*In conclusion*, the combined use of a highly-enantioselective enzyme-catalysed acetylation of a double-meso-shaped tetrakis-hydroxymethyl sulfide and of *E*-selective RBR conditions permitted an efficient conversion of stigmaterol into the more elaborate branched-chain steroid β-4c, an effective precursor of brassinolide.

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## References and Notes

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4- Lee, G. H.; Choi, E. B.; Lee, E.; Pak, C. S. *Tetrahedron Lett.* **1993**, *34*, 4541-4542. The use of NaHg/MeOH induced the formation of compound **17** as a side-product.

5- m.p. 53-55°C; obtained by LAH reduction of the methyl ester of the corresponding carboxylic acid (prepared according to Eliel, E. L.; Banks, H. D. *J. Am. Chem. Soc.* **1972**, *94*, 171-176).

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7- *Selected data:* (*S*)-**10c**:  $[\alpha]_D -4$  (c=8); C, H (%): 73.09, 8.69 (calc.: 73.12, 8.59);  $^1\text{H}$  NMR: 0.83 (d, J=6.9Hz, 3H), 1.07 (s, 9H), 1.96-2.06 (m, 1H); 2.61, s, 1H), 3.56-3.78 (m, 4H); 7.36-7.46 (m, 6H), 7.64-7.75 (m, 4H);  $^{13}\text{C}$  NMR (13.4, 19.3, 27, 37.6, 67.5, 68.6, 127.9, 129.9, 133.3, 135.7; (*R*)-**12c**:  $[\alpha]_D -11$  (c=7);  $^1\text{H}$  NMR: 1.07 (d, 3H, J=6.7Hz), 1.12 (s, 9H), 1.97-2.07 (m, 1H), 2.72-2.82 (dd, J=7.6, 13Hz), 3.28-3.38 (dd, J=5.5, 13Hz, 1H), 3.57-3.65 (dd, J=5.7, 10Hz), 3.69-3.76 (dd, J=5, 10Hz), 7.14-7.51 (m, 11H), 7.69-7.74 (m, 4H) (adding Eu(hfc)<sub>3</sub> to the NMR tube failed to reveal the presence of the corresponding enantiomer);  $^{13}\text{C}$  NMR: 16.5, 19.5, 27, 35.9, 37.2, 67.6, 125.6, 127.8, 128.9, 129.8, 133.8, 135.7, 137.4; (*R*)-**13c**:  $[\alpha]_D -16$  (c=5); C, H (%): 68.99, 7.2 (calc.: 68.98, 7.12);  $^1\text{H}$  NMR: 1.02 (s, 9H), 1.08 (d, J=6.8Hz, 3H); 2.22-2.26 (m, 1H), 2.85-2.96 (dd, J=8.5, 14.2Hz, 1H), 3.38-3.51 (m, 2H), 3.56-3.63 (dd, J=4.4, 10.2Hz, 1H), 7.14-7.51 (m, 13H), 7.69-7.74 (m, 2H); 16.7, 19.3, 26.9, 27, 31.8, 59.2, 67.4, 127.8, 128, 129.4, 129.9, 133.3, 133.6, 135.6, 140.1; (*R*)-**14c**:  $^1\text{H}$  NMR: 1.09 (s, 9H); 1.28 (d, J=7Hz, 3H), 1.31 (s, 3H), 1.37 (s, 3H), 2.22-2.26 (m, 1H), 3.68-3.76 (dd, J=6.1, 10.2Hz, 1H), 3.87-3.94 (dd, J=3.6, 10.2Hz, 1H), 7.35-7.7 (m, 13H), 7.83-7.87 (m, 2H);  $^{13}\text{C}$  NMR: 14, 19.4, 20.1, 20.4, 27, 38.8, 65.9, 66.1, 127.8, 128.8, 129.8, 130.5, 133.4, 133.5, 135.8, 136.4; (*S*)-**15c**:  $[\alpha]_D +3$  (c=3); C, H (%): 77.6, 9.38 (calc.: 77.58, 9.47);  $^1\text{H}$  NMR: 0.85 (d, 3H), 0.93 (d, J=6.8Hz, 6H), 1.13 (s, 9H), 1.55-1.71 (m, 1H), 1.76-1.89 (m, 1H), 3.51-3.59 (dd, J=6.5, 9.9Hz, 1H); 3.62-3.7 (dd, J= 6.1, 9.9Hz, 1H), 7.38-7.52 (m, 6H), 7.7-7.78 (m, 4H);  $^{13}\text{C}$  NMR: 12.9, 18.3, 19.5, 20.9, 27.1, 28.9, 41.6, 67.5, 127.7, 129.6, 134.3, 135.8; (*S*)-**16c**:  $[\alpha]_D +5$  (c=3);  $^1\text{H}$  NMR: 0.81 (d, J=6.8Hz, 3H), 0.83 (d, J=6.9Hz, 3H), 0.88 (d, J=6.8Hz, 3H), 1.41-1.53 (m, 1H), 1.6-1.73 (m, 1H), 1.82 (s, 1H (OH)), 3.36-3.44 (dd, J=6.9, 10.5Hz, 1H), 3.52-3.6 (dd, J=5.9, 10.5Hz, 1H);  $^{13}\text{C}$  NMR: 12.6, 18, 20.7, 28.7, 41.4, 66.6; (*S*)-**8c**:  $[\alpha]_D +2$  (c=7);  $^1\text{H}$  NMR: 0.86 (d, J=6.7Hz, 3H), 0.9 (d, J=6.8Hz, 3H), 0.96 (d, J=6.6Hz, 3H), 1.31-1.41 (m, 1H), 1.59-1.72 (m, 1H), 3.11-3.2 (dd, J=6.9, 9.6Hz, 1H), 3.23-3.3 (dd, J=5, 9.6Hz, 1H);  $^{13}\text{C}$  NMR: 14.1, 16.1, 18.1, 20.5, 32.1, 41.1; **18**: m.p. 51.5-53°C; C, H (%): 44.64, 8.53 (calc.: 45.69, 8.62);  $^1\text{H}$  NMR (CD<sub>3</sub>OD): 1.76-1.89 (m, 2H), 2.58 (d, J=6.8Hz, 4H), 3.63 (d, J=5.6Hz, 8H), 4.87 (m, 4H (OH));  $^{13}\text{C}$  NMR: 32.2, 45, 62.9; (*R,R*)-**22**:  $[\alpha]_D +12$  (c=5); C, H (%): 53.01, 8.16 (calc.: 53.22, 8.12);  $^1\text{H}$  NMR: 1.35 (t, J=7Hz, 3H), 1.88-2.03 (m, 2H), 2.07 (s, 6H), 3.57-3.62 (m, 4H), 4.02-4.17 (m, 4H);  $^{13}\text{C}$  NMR: 20.9, 26.1, 37.7, 62.3, 64.8, 171.4; (*S,S*)-**23**:  $[\alpha]_D -2$  (c=6); C, H (%): 73.7, 8.64 (calc.: 73.84, 8.56);  $^1\text{H}$  NMR: 0.87 (t, J=7.2Hz, 6H), 1.08 (s, 18H), 1.43-1.61 (m, 4H), 1.63-1.69 (m, 2H), 2.55 (dd, J=6, 9.6Hz, 2H), 3.65 (dd, J=5.2, 10Hz, 2H), 3.73 (dd, J=4.6, 10Hz, 2H), 7.34-7.47 (m, 12H), 7.65-7.73 (m, 8H);  $^{13}\text{C}$  NMR: 11.5, 19.4, 23.3, 27, 34.8, 42.4, 64.8, 127.7, 129.7, 133.9, 135.8; (*R,R*)-**24**: C, H (%): 77.82, 8.61 (calc.: 77.72, 8.69);  $^1\text{H}$  NMR: 0.87 (t, J=7.5Hz, 6H), 1.05 (s, 18H), 1.23-1.37 (m, 4H), 1.61-1.74 (m, 2H), 3.49-3.77 (m, 4H), 5.24-5.3 (m, 2H); 7.3-7.45 (m, 12H), 7.65-7.72 (m, 8H); (*R*)-**10a**:  $[\alpha]_D +15$  (c=2); C, H (%): 73.49, 8.84 (calc.: 73.63, 8.82);  $^1\text{H}$  NMR: 0.86 (t, J=6.4Hz, 3H), 1.08 (s, 9H), 1.19-1.33 (m, 2H), 1.65-1.76 (m, 1H), 2.68 (s, 1H (OH)), 3.62-3.86 (m, 4H), 7.37-7.5 (m, 6H), 7.68-7.72 (m, 4H);  $^{13}\text{C}$  NMR: 11.8, 19.3, 20.7, 27, 44.1, 65.9, 67.1, 127.7, 129.9, 133.2, 135.7; (*S*)-**12a**:  $^1\text{H}$  NMR: 0.85 (t, J=7.3Hz, 3H), 1.07 (s, 9H), 1.47-1.58 (m, 2H), 1.7-1.77 (m, 1H), 2.93 (dd, J=6.4, 12.8Hz, 1H), 3.63 (dd, J=5.8, 10.1Hz, 1H), 3.78 (dd, J=4.3, 10.1Hz, 1H), 7.15-7.49 (m, 11H), 7.66-7.71 (m, 4H);  $^{13}\text{C}$  NMR: 11.4, 19.4, 24, 27, 35.2, 42.2, 64.6, 125.5, 127.7, 128.7, 128.8, 129.7, 133.7, 135.7, 137.5; (*R*)-**16a**:  $[\alpha]_D +9$  (c=2);  $^{13}\text{C}$  NMR: 12.2, 19.3, 19.8, 20.3, 27.6, 48.3, 63.1;  $\beta$ -**3c**:  $[\alpha]_D +80$  (c=5); C, H (%): 77.99, 11.16 (calc.: 77.96, 11.28);  $^{13}\text{C}$  NMR: 12.4, 13.2, 15.3, 18, 18.4, 18.9, 20.4, 21.5, 22.6, 24.3, 25, 28.3, 30.6, 31.5, 33.4, 35.1, 35.3, 36.8, 38.3, 39, 40.2, 43.1, 43.4, 48.1, 55.7, 56.4, 56.6, 82.4;  $\beta$ -**4c**:  $[\alpha]_D +42$  (c=4); C, H (%): 84.3, 11.78 (calc.: 84.4, 11.72);  $^{13}\text{C}$  NMR: 12.5, 13.2, 18.2, 19.4, 19.7, 20.3, 21.1, 21.6, 22.9, 24.3, 25.1, 29, 30.6, 33.3, 33.5, 35.2, 35.4, 40.3, 40.4, 42.8, 43.2, 43.5, 48.2, 56.2, 56.7, 57.7, 131.9, 136.2. Excepted as otherwise stated,  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded at 200 and 50MHz, respectively, on CDCl<sub>3</sub> solutions. All  $[\alpha]_D$  refer to CH<sub>2</sub>Cl<sub>2</sub> solutions and were measured at 21°C

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